

M. V. Vasin [2012] Med. Hypotheses Res. 8: 39-49.

The Non-uniformity of the Absorption of Ionizing Radiation Energy in the Body Potentiates the Radioprotective Effects of Drugs: Influence on the Post-radiation Recovery of Radiosensitive Tissues with High-Dose Total-Body Irradiation

Mikhail V. Vasin*

Department of Medicine of Catastrophe, Russian Medical Academy of Post-Graduate Education of Department of Health Care of Russian Federation, Moscow, Russia¹

Abstract. Purpose: This review proposes a hypothesis of the mechanism of potentiating the radioprotector effect with non-uniform irradiation. Conclusion: Preventive application of radioprotectors provides a radioprotective effect not only by means of partially neutralising the "oxygen effect" but also by their pharmacological action on mesenchymal and hematopoietic stem cell progenitors. The stem cells that survive at the site of shielding in the irradiated body may become an object for the radioprotector's action via the stimulation of stem cell proliferation and expansion. Shielding radiosensitive tissues increases the spectrum of radioprotector action and is the basis of the potentiation of combined radioprotector + shielding protection. Significant non-uniformity of the absorption of ionizing radiation energy in a body increases the effectiveness of radioprotective drugs with lethal and extra-lethal levels of irradiation, which must be considered when providing emergency healthcare to individuals exposed to radiation accidents.

* **Correspondence:** Dr. Michail V. Vasin, Department of Medicine of Catastrophe, Russian Medical Academy of Post-Graduate Education, 125284, Moscow, St. Polikarpova 10, Russia. E-mail: mikhail-v-vasin@yandex.ru

Received on 4-1-2012; accepted on 6-7-2012.

1. Introduction

The distribution of the absorbed energy of ionizing radiation through the body of animals and humans is characterized by non-uniformity that increases with an increase in body weight [1]. The maximum absorption of ^{60}Co and ^{137}Cs γ radiation energy occurs at a depth of ~ 10 mm from the surface of the body [2] and shifts to a larger depth with an increase in ionizing radiation energy [3]. The screening of potential radioprotective compounds has been performed in mice, an experimental model with a low body weight that provides a relatively uniform distribution of γ radiation throughout the body. Sixty years ago, Bacq et al. [4] discovered the first radioprotector, cysteamine, in mice.

However, the clear non-uniformity of absorption of γ radiation energy through an animal's body is observed when radioprotector studies are performed on large animals, such as dogs and monkeys. Moreover, because of the greater mass of the human body, this effect should be considered when data from radiobiological experiments are extrapolated from animals to humans who have been exposed to lethal doses of acute radiation during a nuclear accident.

2. Local Shielding, the Abscopal Effect, and Stem Cell Migration: History and the Present

For years, the partial shielding of individual animal body parts has been used as a model of the influence of the non-uniform distribution of ionizing radiation through an animal's body on the course of acute radiation sickness. Investigations in this field included studies by Jacobson and coworkers [5-9] on mice that found a protective effect of shielding the spleen, liver lobe, intestinal loop, and hind limb. All of the cases observed an accelerated postradiation recovery of the hemopoietic system. Shielding of the spleen provided 55% survival of the animals exposed to lethal doses of radiation, indicating

the role of the spleen in these processes. These findings allowed the authors [9] to hypothesize that the observed effect might be caused by the impact of a humoral factor (or factors) secreted by the reticuloendothelial tissue contained in the shielded tissues. Subsequent time-dependent removal of the spleen performed after exposure of the animals to radiation showed that the influence of this individual organ began 1 h later, with a maximal effect observed 24 h after radiation exposure.

Maisin and coworkers [10-15] confirmed a protective effect of shielding the spleen, intestine, liver, and bone marrow in rat studies and found that the liver played an essential role in the increase in the radioresistance of the body. The same observation was made by Gershon-Cohen et al. [16]. Additionally, early investigations by Craves [17] and Edelman [18] showed that shielding the adrenal glands mitigated radiation toxicity. A significant radiation-protective effect was observed with shielding of the abdomen from exposure to high-energy γ -neutron and proton radiation [19-25]. Further studies confirmed a significant impact of locally shielding small sections of the bone marrow on the course of acute radiation syndrome both in small and large animals (dogs) [26-33].

The first evidence of a distant (abscopal) effect was discovered by Barakina [34] and confirmed by more recent studies [35-37]. They reported accelerated postradiation recovery of hemopoietic tissue in intestine-shielded mice exposed to lethal doses of whole-body irradiation. These data were based on the quantitative analysis of bone marrow cellularity, the accelerated elimination of chromosomal aberrations in bone marrow, and an increase in the number of hemopoietic stem cells in the spleen of the animals.

Nonetheless, a reduction of the radiation dose delivered to the shielded intestine at lethal doses of whole-body radiation partially or completely eliminated the intestinal syndrome and toxemic radiation component of acute

radiation sickness and may mitigate the development of bone marrow syndrome. Our studies [38] fully confirmed the existence of the abscopal effect in studies of guinea pigs and rats exposed to non-lethal low-dose γ irradiation (1 and 2 Gy, respectively). Shielding the upper third of the abdomen of animals accelerated the elimination of chromosomal aberrations in bone marrow to levels equivalent to a two-fold reduction of the radiation dose. The effect was registered 2-3 days after irradiation.

The abscopal effect was also manifested as an acceleration of dermal injury healing in animals exposed to whole-body irradiation with small shielded sections of bone marrow [39]. Hanks [40] was the first to suggest that a shielded area of bone marrow might be a source of endogenous colony-forming units in the spleen (CFUs) in irradiated mice, although no direct data that support this possibility was yet provided. The emergence of CFUs after partially shielding the bone marrow in irradiated animals was confirmed [41, 42]. Petrov et al. [43] obtained convincing evidence of stem cell migration from the bone marrow shielding site to further the process of cell repopulation. They used a gene marker in experiments in two different inbred irradiated parabiotic mice, in which one had shielding of the paw. The time necessary for completing hematopoietic cell expansion from the bone marrow shielding through the entire hematopoietic tissue of the irradiated body was 6 and 14 days in rats and dogs, respectively, and allowed the survival of the animals after lethal radiation exposure [44, 45].

Nothdurft et al. [46,47] performed a similar study in dogs exposed to radiation of the upper half of the body at a dose of 11.7 Gy, resulting in the loss of 70% of the bone marrow, and discovered the concurrent activation of the hematopoietic system in the non-irradiated part of the body and emergence of granulocyte/macrophage (GM) progenitor cells in the blood flow and irradiated segment of hematopoietic tissue 1 week after irradiation.

Under the same experiment conditions, they also discovered that recombinant human granulocyte colony-stimulating factor (rhG-CSF) application increased the number of GM progenitor cells in the blood flow and significantly accelerated the hematopoietic recovery of the irradiated sites of the body, particularly within the first 7 days after exposure [48].

The most significant parts of the body for shielding against radiation were established in experiments that used equal weights and absorbed radiation doses of shielded body parts in dogs and rats [22-25]. The greatest protective effect, with a dose reduction factor (DRF) of 2.4, was found in the animals exposed to lethal doses of irradiation when the upper half or third of their abdomen was shielded [23]. A mathematical model that evaluates the effectiveness of the partial local shielding of the abdomen as a function of the width and thickness of a shield and absorbed dose has been developed [49]. Shielding the upper parts of the abdomen protected the liver, spleen, adrenal glands, one third of the intestine, and a small portion of the bone marrow in four vertebrae and partially in the lower ribs. Notably, this is an area where a large part of the reticuloendothelial system of the body (e.g., liver, spleen) is located. Lowering radiation exposure in these tissues to sub-lethal doses by means of partial shielding is likely to generate active stromal foci that are essential for the post-radiation restoration of hematopoiesis in animals exposed to lethal doses of whole-body irradiation [50-52]. Additionally, partially shielding the abdomen reduces the development of post-exposure primary radiation syndrome [53], namely primary hypercorticism that suppresses stem cell migration [54-58]. Lowering radiation exposure in the adrenal glands by shielding the abdomen may be favourable for the mitigation of acute radiation syndrome [17, 18, 59].

Intensive and comprehensive investigations of the biochemical and cellular mechanisms of hematopoietic stem cell (HSC) traffic from the bone marrow to blood flow and from the

circulation to stem cell niches to continue hematopoiesis at new sites have been performed over the past decade [60]. Although many issues have not yet been resolved, some common features of these processes are discussed below. First, anatomical microsites of the bone marrow where HSCs are preserved and reproduced are niches with a complex three-dimensional structure of the bone marrow microenvironment, consisting of stromal cells and capillary endothelia. The niche regulates the adequate level of hematopoiesis through cytokine production (G-CSF, stem cell factor, interleukin-6, leukaemia inhibitory factor, and so on), thereby providing long-term HSC proliferation, especially the proliferation of the cells responsible for myelopoiesis and thrombocytopoiesis. Niches also work as a barrier to adjust hematopoietic stem and progenitor cell (HSPC) traffic and homing. Reciprocal SDF-1/CXCR4 interactions between hematopoietic and bone marrow stromal cells play a crucial role in HSPC retention in the niche. Chemokine SDF-1 is produced in the bone marrow, spleen, and other tissues exclusively by endothelial and stromal cells. The level of SDF-1 concentration and expression of the CXCR4 receptor determine HSPC and mesenchymal stem cell (MSC) homing and proliferation [61, 62]. G-CSF also plays an important role as the major factor that mobilizes stem cells. G-CSF creates beneficial conditions for the egress of HSPC from the bone marrow by decreasing CXCR4 expression in HSPC [63, 64].

The physiological importance of HSPC and MSC migration through repopulation is clearly associated with the repair of radiation-induced hematopoietic system damage [65, 66]. Irradiation increases the expression of HSPC mobilization factors, including G-CSF [67-70]. The activation of reticuloendothelial tissues induced by radiation is the most important for these processes [50-52]. A study by Francois et al. [71] provided evidence that whole-body irradiation increases the homing of intravenously injected human MSC in the tissues

of nude (-/-) mice. MSCs support the expansion of HSCs after irradiation [72, 73]. In another study, the transplantation of irradiated CD34 (+) cells together with MSC in irradiated baboons greatly rescued CD34+ cells from radiation-induced apoptosis and enhanced the therapeutic effect of CD34 (+) cells [74]. Therefore, the beneficial effect of preliminary whole-body irradiation on the repair of local radiation-induced skin injury and diabetic wounds can be explained by the increased migration of mesenchymal stem cells [75, 76].

Overall, the reduction of radiation absorption up to sub-lethal doses by shielding individual parts of the body during whole-body irradiation preserves a certain number of viable hematopoietic or mesenchymal stem cells that further migrate through the body under the influence of radiation-induced humoral factors, thereby providing conditions for the repair of damaged tissues.

3. Potentiation of the radioprotective effect of the combined application of local shielding and radioprotectors

Presently, the pharmacological classification of radioprotective agents reflects the most significant historical advances in the screening of various chemical compounds and theoretical knowledge of their radioprotective mechanism. With the exception of pharmaceutical countermeasures against radioactive nuclide incorporation or primary radiation syndrome, radioprotective agents can be referred to as the following: (1) radioprotectors ("chemical protection" by Bacq) that exert their effects on physicochemical and biochemical levels in cells during exposure to ionizing radiation, (2) radiomitigators (e.g., oestrogens, androgens, cytokines, and immunological adjuvants) that promote the acceleration of the post-radiation restoration of myelopoiesis by stimulating HSPC and MSC mobilization, and (3) radiomodulators (e.g., vitamins and nutritional supplements) that increase the resistance of the body to irradiation

and other unfavourable environmental factors by adaptively shifting the effectiveness of the antioxidative protection of the organism [77].

The mechanism of the radioprotector effect is based on the realization of the "oxygen effect," a radiobiological phenomenon. Radioprotectors either induce lower oxygen content in cells by means of pharmacological action via specific receptors (e.g., serotonin, mexamine, indralin) or directly provide thiol groups (i.e., sulphur-containing radioprotectors, such as cysteamine and amifostine) in competitive chemical reactions with oxygen for products from DNA radiolysis [78, 79].

The first data on the combined application of local shielding and radioprotective agents were reported by Jacobson [9]. He noted an additive protective effect of both cysteine (or estradiol) and the shielding of the spleen in mice exposed to lethal doses of radiation. The first investigation of combined protection that applied both shielding and the radioprotective drug cysteamine was performed by Maisin and coworkers [80, 81]. They discovered a synergistic effect of shielding the liver and intraperitoneal cysteamine administration in rats exposed to lethal doses of radiation. Saxonov and coworkers [82, 83] later estimated the efficiency of combined protection for various groups of radioprotectors and shielding various parts of animals' bodies. In studies of rats exposed to lethal doses of 10 Gy whole-body γ irradiation, partially shielding the abdomen or head provided 30% survival of the animals. Under this condition, cysteamine or mexamine at non-protective doses potentiated the radioprotective effect of shielding, with up to 80% survival of the animals [82]. The potentiation and synergism of the protective action of the combined application of radioprotectors and shielding of radiosensitive tissue have been repeatedly confirmed by other investigations [84-87].

To extrapolate these data to humans, experiments on large animals have been performed. Clearly marked non-uniform absorption of ionizing radiation energy in the

animal's body must be considered. Partially shielding both the abdomen and head and administering the emergency radioprotector indralin pre-irradiation revealed a strongly pronounced effect of the combined protection in studies on irradiated dogs. With 10-12 Gy whole-body γ irradiation, a dose that is 2.5- to 3-fold higher than the lethal dose of irradiation for this species, 75% of the animals with combined protection survived compared with 100% mortality in the experimental groups that received either the shielding or radioprotector alone [88].

Furthermore, the potentiating effect of bone marrow (pelvis) shielding together with indralin administration was confirmed in experiments on dogs exposed to 5 Gy high-energy proton (~240 M α B) irradiation [89]. The absorbed radiation dose in all of the experimental groups was identical. Under these conditions, the combined protection provided 57% survival of the dogs, compared with 100% mortality in the experimental groups that received either shielding or indralin alone.

4. Plausible mechanism of the potentiating effect of combined protection

A possible mechanism of the observed potentiation of the effect of combined protection is based on the concept of augmenting the reproduction and migration of MSC and HSPC from the shielding site under the influence of the pharmacological action of radioprotectors, causing their further expansion to radiation-damaged hematopoietic tissues. Therefore, the influence of radioprotectors on these processes may be detected if radioprotectors are applied after radiation exposure, when they fail to exert a protective effect as antagonists of the "oxygen effect" [77].

Rixon et al. [90] first revealed the therapeutic properties of serotonin repeatedly injected in rats 1 h after irradiation. This effect was later confirmed not only with receptor-mediated radioprotectors (e.g., serotonin,

mexamine, and indralin) but also with sulphur-containing radioprotectors (e.g., cystamine and AET) [91-93]. With liver shielding in rats, Masin et al. [80, 81] found a protective effect of cysteamine administered after whole-body lethal-dose irradiation when the radioprotector alone was ineffective. In our studies in rats exposed to 10 Gy radiation with shielding of the upper quarter of the abdomen, including protection of the liver, indralin administered after irradiation increased survival by as much as 55%, whereas the radioprotector alone had no protective effects [94]. Indralin applied after whole-body irradiation also increased the amount of CFUs in mice with paw shielding [88]. The therapeutic effects of radioprotectors have been confirmed in large animals. Indralin used within 30 min after whole-body absolutely lethal irradiation increased survival in irradiated dogs by as much as 30% and 60% when applied alone and coupled with acute radiation sickness therapy accordingly [88, 95]. In this condition preventive application of indralin provided 90% animal survival, in contrast to the therapeutic post-exposure effect [88].

Orbeli [96], an adherent of Pavlov, was the first to discover the adaptation-trophic role of the sympathetic nervous system. Adrenergic agents and serotonin were later found to stimulate hemopoiesis and stem cell mobilization and migration [60, 97-101]. Moreover, norepinephrine, mexamine (i.e., the 5-methyl-derivate of serotonin), and indralin as a direct α_1 -adrenergic agent [102] significantly reduced the hemotoxicity of carboplatin [103-105]. The proposed direct action of these tested agents on the hematopoietic system was confirmed by recent discoveries of α_1 -, α_2 -, β_2 -adrenoceptors and serotonin 5-HT₂ receptors on HSPC [106, 107].

Indralin was also shown to activate ribonucleotide reductase in the bone marrow and spleen in "viable animals," which catalyses the inclusion of ribonucleotide-5-diphosphate in DNA through the synthesis of desoxyribonucleotide triphosphate [108-110]. A

similar action of norepinephrine on nucleotide metabolism was shown in bone marrow MSC by increasing cell DNA synthesis through α_1 -adrenergic receptors [111]. A stimulatory effect of adrenergic agents and serotonin on hematopoietic tissue is likely realized through the hematopoietic microenvironment by means of stromal cell activation. The therapeutic effect of sulphur-containing radioprotectors is likely to be indirectly initiated by the stimulation of the adrenergic system through its vasodilative action [112] and carried out by mesenchymal cells [113].

5. Conclusion

The stem cells that survive at the site of shield protection of the irradiated body may become the object for the action of radioprotectors via stimulation of MSC and HSC proliferation and expansion. Therefore, without shielding, radioprotectors have only therapeutic effects in small animals that are exposed to whole-body \sim LD₇₀₋₈₀ acute irradiation and consistently demonstrate a lack of effectiveness at lethal radiation doses [93] that likely result in the complete radiation-induced damage of the substrate of radioprotector action (i.e. HSPC). A more favourable scenario can be observed with irradiation in large animals (e.g. dogs) [88] and may be extrapolated to humans because of their larger size, greater heterogeneity, and non-uniformity of absorption of ionizing radiation energy.

The preventive application of radioprotectors is likely to provide a radioprotective effect not only by partially neutralizing the "oxygen effect" but also by their pharmacological stimulatory action on mesenchymal stem and hematopoietic stem cell progenitors. Shielding radiosensitive tissues increases the spectrum of radioprotector action and is the basis of combined protection. The significant non-uniformity of the absorption of ionizing radiation energy in a body increases the effectiveness of radioprotective drugs to extra-

lethal levels of irradiation and should be considered when providing emergency healthcare to individuals exposed to radiation accidents.

References

- [1] **Baltschukat K and Nothdurft W** [1990] Hematological effects of unilateral and bilateral exposures of dogs to 300-kVp X rays. *Radiat Res* 123: 7-16.
- [2] **IAEA** [1999] Safety standards series. Assessment of occupational exposure due to sources of radiation. Safety guide. RS-G-1.3. Vienna: Atomic Energy Agency.
- [3] **Smiljanic I and Schultz FW** [2004] Applicability and limitations of the Adam mathematical phantom with respect to radiological protection. *Radiat Prot Dosimetry* 11: 89-92.
- [4] **Bacq ZM, Herve A, Lecomte J, Fischer P and Blavier J** [1951] Protection contre le rayonnement X par la beta-mercaptoethylamine. *Arch Int Physiol* 59: 442-447.
- [5] **Jacobson LO, Marks EK, Gaston E, Robson M and Zirkle R** [1949] The role of the spleen in radiation injury. *Proc Soc Exptl Biol Med* 70: 740-742.
- [6] **Jacobson LO, Simmons EL, Marks EK, Robson MJ, Bethyard WF and Gaston EO** [1950] The role of the spleen in radiation injury and recovery. *J Lab Clin Med* 35: 746-770.
- [7] **Jacobson LO, Simmons EL, Marks EK and Eldredge JH** [1951] Recovery from radiation injury. *Science* 113: 510-511.
- [8] **Jacobson LO, Simmons EL, Marks EK, Gaston EO, Robson MJ and Eldredge JH** [1951] Further studies on recovery from radiation injury. *J Lab Clin Med* 37: 683-697.
- [9] **Jacobson LO** [1952] Evidence for a humoral factor (or factors) concerned in recovery from radiation injury: a review. *Cancer Res* 12: 315-325.
- [10] **Mandart M, Lambert G and Maisin J** [1952] Protection of various parts of the body of rats exposed to a lethal dose of x-rays; influence of the spleen. *C R Soc Biol Fil* 146: 1305-1307.
- [11] **Mandart M, Lambert G, Maisin H. and Maisin J** [1952] Importance of the protection of the liver area in rats subject to a lethal dose of x-rays. *C R Soc Biol Fil* 146: 1647-1649.
- [12] **Maisin J, Dunjic A, Van Lancker J, Lambert G and Passau L** [1953] Effect of hepatic protection on the survival of animals irradiated in toto: importance of protected zone. *C R Soc Biol Fil* 147: 1520-1522.
- [13] **Maisin J, Dunjic A and Maisin H** [1954] Importance of the protection of the liver region in rats exposed to a fatal dose of x-rays: attempted interpretation. *C R Soc Biol Fil* 148: 611-615.
- [14] **Maisin J, Maisin H and Dunjic A** [1954] Role of intestinal and bone marrow synergism in the protection of the rat against fatal dose of x-rays. *C R Soc Biol Fil* 148: 743-745.
- [15] **Maisin JH, Van Lancker J, Lambert G, Passau L, Mandart M, Dunjic A and Maisin H** [1954] The role of the liver region in the protection against ionising radiations. *Acta Radiol Suppl* 116: 40-48.
- [16] **Gershon-Cohen J, Hermel MB and, Griffith JQ** [1951] The value of small lead shields against the injurious effect of total body irradiation. *Science* 114:157-158.
- [17] **Craves BN** [1948] The effect of adrenal cortical injury on the toxicity of roentgen rays. *Am J Roentgenol Rad Ther* 59: 404-407.
- [18] **Edelmann A** [1951] Adrenal shielding and survival of rats after X-irradiation. *Am J Physiol* 165: 57-60.
- [19] **Bond VF, Swift MN, Allen AC and Fishler MC** [1951] Sensitivity of abdomen of rat to x-irradiation. *Am J Physiol* 161: 323-330.
- [20] **Swift MN, Taketa ST and Bond VF** [1954] Effect of partial shielding of rat intestine during x-irradiation. *Fed Proc* 13(1 Part 1): 523.
- [21] **Swift MN and Taketa ST** [1956] Modification of acute intestinal radiation syndrome through shielding. *Am J Physiol* 185: 85-91.
- [22] **Razgovorov BL, Morozov VS, Shashkov VS, Antipov VV, Dobrov NN, Konnova NI, L'vova TS and Saksonov PP** [1965] [The influence of of some of body shielding on reduction of radiation reaction under exposure of γ -rays and proton of high energy]. Russian In: Sisakian NM (Editor), *Problemy Kosmicheskoi Biologii*. Vol. 4. Moscow: Nauka, pp. 411-429.
- [23] **Razgovorov BL and Morozov VS** [1967] [Animal survival after total γ -radiation under shielding of abdomen]. Russian. In: Sisakian NM (Editor), *Problemy Kosmicheskoi Biologii*. Vol. 6. Moscow: Nauka, pp. 448-459.
- [24] **Razgovorov BL** [1971] Influence of shielding certain parts of the body on the course of radiation sickness and survival of animals with total gamma-neutron irradiation. In: Saksonov PP and Davydov BI (Editors), *Radiobiological aspects of the reactivity of organism during space flights. Problems of space biology*. Vol. 14. Moscow: Nauka, English translation accessed 1973 by NASA TT F-721. Washington DC: National Aeronautics and Space Administration, pp. 267-287.
- [25] **Razgovorov BL and Konnova NI** [1971] Influence of shielding certain parts of the body on the course of radiation sickness in dogs with total gamma-irradiation. In: Saksonov PP and Davydov BI (Editors), *Radiobiological aspects of the reactivity of organism during space flights. Problems of space biology*. Vol. 14. Moscow: Nauka, English translation accessed 1973 by NASA TTF-721. Washington DC: National Aeronautics and Space Administration, pp. 307-329.
- [26] **Storer JB, Lushbaugh CC and Furchner JE** [1952] The protective effect of shielded ectopic bone marrow against total body x-radiation. *J Lab Clin Med* 40: 355-366.

- [27] **Lamerton LF, Elson LA and Harriss EB** [1953] A study of the phases of radiation response in the rat. II. The effects of non-uniform irradiation. *Brit J Radiol* 26: 568-576.
- [28] **Loiselle JM, Lamerton LF and Adams K** [1959] Comparison of protective effects of hind-leg shielding and isologous bone marrow injection in x-irradiated rats. *Int J Radiat Biol* 1: 266-276.
- [29] **Osmond DG, Roylance PJ, Lee WR, Ramsell TG and Yoffey JM** [1966] The effect of unilateral limb shielding on the haemopoietic response of the guinea-pig to gamma irradiation (150 r). *Brit J Haematol* 12: 365-375.
- [30] **de Vries FA and Vos O** [1966] Prevention of the bone-marrow syndrome in irradiated mice. A comparison of the results after bone-marrow shielding and bone-marrow inoculation. *Int J Radiat Biol* 11: 235-243.
- [31] **Cole LJ, Haire HM and Alpen EL** [1967] Partial shielding of dogs: effectiveness of small external epicondylar lead cuffs against lethal x-radiation. *Radiat Res* 32: 54-63.
- [32] **Kalandarova MP** [1973] [Comparative characteristics of bone marrow hematopoiesis in dogs following acute irradiation by protons in conditions of partial shielding of a portion of the bone marrow]. *Radiobiologiya* 13: 774-778. Russian
- [33] **Nevskaia GF, Abramova GM, Ginsburg EV, Ishmukhametova DN and Skorik AS** [1977] [Hematopoiesis in dogs irradiated with protons in lethal doses with screening of the bone marrow]. *Kosm Biol Aviakosm Med* 11: 67-70. Russian
- [34] **Barakina NF** [1962] On the favorable effect of screening of nonhematopoietic organs on the restoration of hemopoiesis in irradiated animals. *Radiobiologiya* 2: 42-47.
- [35] **Ianushevskaja MI** [1965] The effect of intestinal screening on the restoration of haemopoiesis in irradiated mice. *Dokl Akad Nauk SSSR* 164: 445-447.
- [36] **Murphy MJ, Porteous DD and Gordon AS** [1967] Hematopoietic repopulation after intestinal shielding from x-irradiation. *Nature* 215: 772-773.
- [37] **Vávrová J and Petýrek P** [1984] Shielding of the abdominal region during X-irradiation: effect on haemopoietic stem cells. *Folia Biol (Praha)* 30: 267-275.
- [38] **Vasin MV and Razgovorov BL** [1971] Effect of shielding stomach on the frequency of chromosome aberration in the cells of marrow of guinea pigs and rats with gamma-irradiation at doses of 50–200 r. In: Saksonov PP and Davydov BI (Editors), *Radiobiological aspects of the reactivity of organism during space flights. Problems of space biology. Vol. 14.* Moscow: Nauka, English translation accessed 1973 by NASA TT F-721. Washington DC: National Aeronautics and Space Administration, pp. 330-338.
- [39] **Stromberg LR, Woodward KT, Mahin DT and Donati RM** [1967] Altered wound healing in x-irradiated rats: the effect of bone marrow shielding. *Experientia* 23: 1064-1065.
- [40] **Hanks GE** [1964] In vivo migration of colony-forming units from shielded bone marrow in the irradiated mouse. *Nature* 203:1393-1395.
- [41] **Robinson CV, Commerford SL and Bateman JL** [1965] Evidence for the presence of stem cells in the tail of the mouse. *Proc Soc Exp Biol Med* 119: 222-226.
- [42] **Croizat H, Frindel E and Tubiana M** [1976] Abscopal effect of irradiation on haemopoietic stem cells of shielded bone marrow-role of migration. *Int J Radiat Biol* 30: 347-358.
- [43] **Petrov RV and Khaitov RM** [1972] [Migration of stem cells from screened bone marrow following non-uniform irradiation]. *Radiobiologiya* 12: 69-76. Russian
- [44] **Vishniakov IuS and Strelin GS** [1979] [Hematopoietic stem cell migration study in dogs]. *Bull Eksp Biol Med* 88: 714-716. Russian
- [45] **Gronskaia NF and Strelin GS** [1979] [Reimmigration of bone marrow stem cells in rats]. *Bull Eksp Biol Med* 87: 589-591. Russian
- [46] **Nothdurft W, Calvo W, Klinnert V, Steinbach KH, Werner C and Fliedner TM** [1986] Acute and long-term alterations in the granulocyte/macrophage progenitor cell (GM-CFC) compartment of dogs after partial body irradiation: irradiation of the upper body with a single myeloablative dose. *Int J Radiat Oncol Biol Phys* 12: 949-957.
- [47] **Nothdurft W and Kreja L** [1998] Hemopoietic progenitor cells in the blood as indicators of the functional status of the bone marrow after total-body and partial-body irradiation: experiences from studies in dogs. *Stem Cells* 16 Suppl 1: 97-111.
- [48] **Nothdurft W, Kreja L and Selig C** [1997] Acceleration of hemopoietic recovery in dogs after extended-field partial-body irradiation by treatment with colony-stimulating factors: rhG-CSF and rhGM-CSF. *Int J Radiat Oncol Biol Phys* 37: 1145-1154.
- [49] **Osokina TF, Davydov BI and Razgovorov BL** [1975] [Prognosis of the survival of rats with partial shielding of the abdominal region] *Radiobiologiya* 15: 784-786. Russian
- [50] **Nowell PC and Cole LJ** [1967] Clonal repopulation in reticular tissues of x-irradiated mice: effect of dose and limb-shielding. *J Cell Physiol* 70: 37-44.
- [51] **Wolf NS and Trentin JJ** [1968] Hemopoietic colony studies. V. Effect of hemopoietic organ stroma on differentiation of pluripotent stem cells. *J Exp Med* 127: 205-214.
- [52] **Sljivić VS** [1970] Radiation and the phagocytic function of the reticuloendothelial system. I. Enhancement of RES function in x-irradiated mice. *Br J Exp Pathol* 51: 130-139.
- [53] **Osokina TF, Razgovorov BL and Davydov BI** [1983] [Prediction of vomiting in dogs after irradiation with shielding of the middle part of the abdomen]. *Kosm Biol Aviakosm Med* 17: 70-72. Russian
- [54] **Khaitov RM, Petrov RV, Moroz BB and Bezin GI** [1974] [Factors controlling recirculation of stem cells. I. Effect of adrenalectomy on the migration of hematopoietic stem cells from the screened bone

- marrow during irradiation]. *Radiobiologia* 14: 516-521. Russian
- [55] **Khaitov RM, Petrov RV, Moroz BB and Bezin GI** [1975] The factors controlling stem cell recirculation. I. Migration of hemopoietic stem cells in adrenalectomized mice. *Blood* 46: 73-77.
- [56] **Bezin GI, Khaitov RM, Moroz BB, Petrov RV and Romashko OO** [1975] The factors controlling stem cell recirculation. II. ACTH-induced inhibition of migration of hemopoietic stem cells. *Blood* 46: 79-84.
- [57] **Bezin GI, Moroz BB, Petrov RV, Khaitov RM and Romashko OO** [1977] [Migration and differentiation of hematopoietic stem cells depending on the level of endogenous corticoids in the body]. *Probl Gematol Pereliv Krovi* 22: 21-27. Russian
- [58] **Moroz BB, Petrov RV, Bezin GI, Khaitov RM and Romashko OO** [1978] [Role of endogenous glucocorticoids in the regulation of migration and recirculation of the hematopoietic stem cells]. *Patol Fiziol Eksp Ter* (5):9-15. Russian
- [59] **Cohen EP, Bruder ED, Cullinan WE, Ziegler D and Raff H** [2011] Effect of high-dose total body irradiation on ACTH, corticosterone, and catecholamines in the rat. *Transl Res* 157: 3847.
- [60] **Papayannopoulou T, Priestley GV, Bonig H and Nakamoto B** [2003] The role of G-protein signaling in hematopoietic stem/progenitor cell mobilization. *Blood* 101: 4739-4747.
- [61] **Dar A, Kollet O and Lapidot T** [2006] Mutual, reciprocal SDF-1/CXCR4 interactions between hematopoietic and bone marrow stromal cells regulate human stem cell migration and development in NOD/SCID chimeric mice. *Exp Hematol* 34: 967-975.
- [62] **Kyriakou C, Rabin N, Pizzey A, Nathwani A and Yong K** [2008] Factors that influence short-term homing of human bone marrow-derived mesenchymal stem cells in a xenogeneic animal model. *Haematol* 93: 1457-1465.
- [63] **Kim HK, De La Luz Sierra M, Williams CK, Gulino AV and Tosato G** [2006] G-CSF down-regulation of CXCR4 expression identified as a mechanism for mobilization of myeloid cells. *Blood* 108: 812-820.
- [64] **Zhang Y, Cheng G, Yang K, Fan R, Xu Z, Chen L, Li Q, Yang A and Jin B** [2009] A novel function of granulocyte colony-stimulating factor in mobilization of human hematopoietic progenitor cells. *Immunol Cell Biol* 87:428-432.
- [65] **Flidner TM, Graessle D, Paulsen C and Reimers K** [2002] Structure and function of bone marrow hemopoiesis: mechanisms of response to ionizing radiation exposure. *Cancer Biother Radiopharm* 17: 405-426.
- [66] **Greenberger JS and Epperly M** [2009] Bone marrow-derived stem cells and radiation response. *Semin Radiat Oncol* 19: 133-139.
- [67] **Peterson VM, Adamovicz JJ, Elliott TB, Moore MM, Madonna GS, Jackson WE 3rd, Ledney GD and Gause WC** [1994] Gene expression of hematoregulatory cytokines is elevated endogenously after sublethal gamma irradiation and is differentially enhanced by therapeutic administration of biologic response modifiers. *J Immunol* 153: 2321-2330.
- [68] **Gaugler MH, Squiban C, Mouthon MA, Gourmelon P and van der Meeren A** [2001] Irradiation enhances the support of haemopoietic cell transmigration, proliferation and differentiation by endothelial cells. *Brit J Haematol* 113: 940-950.
- [69] **Xiao M, Inal CE, Parekh VI, Li XH and Whitnall MH** [2009] Role of NF-kappaB in hematopoietic niche function of osteoblasts after radiation injury. *Exp Hematol* 37: 52-64.
- [70] **Cho KA, Joo SY, Han HS, Ryu KH and Woo SY** [2010] Osteoclast activation by receptor activator of NF-kappaB ligand enhances the mobilization of hematopoietic progenitor cells from the bone marrow in acute injury. *Int J Mol Med* 26: 557-563.
- [71] **François S, Bensidhoum M, Moussedine M, Mazurier C, Allenet B, Semont A, Frick J, Saché A, Bouchet S, Thierry D, Gourmelon P, Gorin NC and Chapel A** [2006] Local irradiation not only induces homing of human mesenchymal stem cells at exposed sites but promotes their widespread engraftment to multiple organs: a study of their quantitative distribution after irradiation damage. *Stem Cells* 24: 1020-1029.
- [72] **Werts ED, Gibson DP, Knapp SA and DeGowin RL** [1980] Stromal cell migration precedes hemopoietic repopulation of the bone marrow after irradiation. *Radiat Res* 81:20-30.
- [73] **Mourcin F, Grenier N, Mayol JF, Lataillade JJ, Sotto JJ, Hérodin F and Drouet M** [2005] Mesenchymal stem cells support expansion of in vitro irradiated CD34(+) cells in the presence of SCF, FLT3 ligand, TPO and IL3: potential application to autologous cell therapy in accidentally irradiated victims. *Radiat Res* 164: 1-9.
- [74] **Drouet M, Mourcin F, Grenier N, Delaunay C, Mayol JF, Lataillade JJ, Peinnequin A and Hérodin F** [2005] Mesenchymal stem cells rescue CD34+ cells from radiation-induced apoptosis and sustain hematopoietic reconstitution after coculture and cointegration in lethally irradiated baboons: is autologous stem cell therapy in nuclear accident settings hype or reality? *Bone Marrow Transplant* 35: 1201-1209.
- [75] **Rezvani M, Ross GA, Wilkinson JH and Bywaters A** [2002] Evidence for humoral effects on the radiation response of rat foot skin. *Br J Radiol* 75: 50-55.
- [76] **Guo WY, Wang GJ, Wang P, Chen Q, Tan Y and Cai L** [2010] Acceleration of diabetic wound healing by low-dose radiation is associated with peripheral mobilization of bone marrow stem cells. *Radiat Res* 174: 467-479.
- [77] **Vasin MV** [2006] [Medicine of prophylaxis and treatment of radiation injury]. Moscow: Medical Academy of Post-Graduate Education. Russian
- [78] **Zheng S, Newton GL, Ward JF and Fahey RC** [1992] Aerobic radioprotection of pBR322 by thiols: effect of

- thiol net charge upon scavenging of hydroxyl radicals and repair of DNA radicals. *Radiat Res* 130:183-193.
- [79] **Savoie C, Swenberg C, Hugot S, Sy D, Sabbattier R, Charlier M and Spothem-Maurizot M** [1997] Thiol WR-1065 and disulphide WR-33278, two metabolites of the drug ethylol (WR-2721), protect DNA against fast neutron-induced strand breakage. *Int J Radiat Biol* 71:193-202.
- [80] **Maisin J, Mandart M, Lambert G and Maisin H** [1953] Curative action of beta-mercaptoethylamine in the rat irradiated with the liver protected. *C R Soc Biol Fil* 147: 362-364.
- [81] **Maisin JH, Lambert G, Mandart M and Maisin H** [1953] Therapeutic action of glutathione and beta-mercaptoethylamine against a lethal dose of x-rays. *Nature* 171: 971.
- [82] **Razgovorov BL, Saksonov PP, Antipov VV, Shashkov VS and Morozov VS** [1971] Change of reactivity of animal to certain pharmacological preparations with parts of the body shielded during total irradiation. In: Saksonov PP and Davydov BI (Editors), *Radiobiological aspects of the reactivity of organism during space flights. Problems of space biology*. Vol. 14. Moscow: Nauka, English translation accessed 1973 by NASA TT F-721. Washington DC: National Aeronautics and Space Administration, pp. 288-306.
- [83] **Saxonov PP** [1975] Protection against radiation (biological, pharmacological, chemical, physical). *Foundation of Space Biology and Medicine*. 3, Washington: NASA, pp. 316-347.
- [84] **Barkaia VS** [1967] [The increase of protection effect of bone marrow shielding by chemical radioprotector under acute radiation injury]. Russian. In: *Lapin BA* (Editor), *Medistinskaiia Primatologiia*. Tbilisi. pp 359-363.
- [85] **Minkova M and Baldzhijska M** [1989] Testing a combined radiation protection modality: chemical protector and local shielding. *Radiobiol Radiother* (Berlin) 30: 277-280.
- [86] **Shashkov VS, Karsanova SK and Iasnetsov VV** [2007] [Comparative protective action of radioprotectors and shielding in gamma-irradiated mice]. *Aviakosm Ekol Med* 41: 39-43. Russian
- [87] **Shashkov VS, Karsanova SK and Iasnetsov VV** [2008] [Protective action of radioprotectors and shielding against high-energy protons in experiments with rats]. *Aviakosm Ekol Med* 42: 58-61. Russian
- [88] **Ilyin LA, Rudnyi NM, Suvorov NN, Chernov GA, Antipov VV, Vasin MV, Davydov BI and Mikhailov PP** [1994] [Indralin – radioprotector of urgent action: radiation-protective property, pharmacology, mechanism of action, clinics]. Moscow: Ministry of Health Care of Russian Federation. Russian
- [89] **Shashkov VS, Efimov VI, Vasin MV, Antipov VV, Iliukhin AV, Vlasov PA, Karsanova SK, Grigor'ev IuG and Ushakov IB** [2010] [Indralin – a novel effective radioprotector during irradiation by high energy protons]. *Aviakosm Ekol Med* 44: 15-20. Russian
- [90] **Rixon EH and Baird KM** [1968] The therapeutic effect of serotonin on the survival of x-irradiated rats. *Radiat Res* 33: 395-402.
- [91] **Shashkov VS, Anashkin OD, Suvorov NN and Manaeva IA** [1971] [Efficiency of serotonin, AET and cystamine at repeated injection after irradiation]. *Radiobiologiia* 11: 621-623. Russian
- [92] **Graevskii EIa, Ianushevskaiia MI, Bueverova EI, Bragina EV and Konstantinova MM** [1981] [Radioprotective activity and aspects of the mechanism of action of biogenic amines in vitro on cultured mammalian cells]. *Radiobiologiia* 21: 683-687. Russian
- [93] **Vasin MV, Ushakov IB, Kovtun VIu, Komarova SN, Semenova LA, Koroleva LV, Galkin AA and Afanas'ev RV** [2008] [The characteristic of radioprotective properties of a radioprotectant B-190 at its administration after radiation]. *Radiats Biol Radioecol* 48: 730-733. Russian
- [94] **Vasin MV, Ushakov IB, Kovtun VIu, Komarova SN, Semenova LA, Galkin AA and Afanas'ev RV** [2008] [Radioprotective properties of a radioprotector of emergency action indraline at its administration after irradiation in conditions of local shielding of a rat abdomen.] *Radiats Biol Radioecol* 48: 199-202. Russian
- [95] **Kolesnichenko IS, Mikhailov LS, Boiarinov AS and Grishin AV** [2005] [Radiation protective scheme of prophylaxis and treatment of official dogs]. *Veterinariia*. (12): 52-54. Russian
- [96] **Orbeli LA** [1949] Adaptation-trophic role of the sympathetic nervous system and of the cerebellum. *Fiziol Zh SSSR im IM Sechenova* 35: 594-595.
- [97] **Papayannopoulou T and Scadden DT** [2008] Stem-cell ecology and stem cells in motion. *Blood* 111: 3923-3930.
- [98] **Katayama Y, Battista M, Kao WM, Hidalgo A, Peired AJ, Thomas SA and Frenette PS** [2006] Signals from the sympathetic nervous system regulate hematopoietic stem cell egress from bone marrow. *Cell* 124: 407-421.
- [99] **Spiegel A, Shivtiel S, Kalinkovich A, Ludin A, Netzer N, Goichberg P, Resnick I, Hardan I, Ben-Hur H, Nagler A, Rubinstein M and Lapidot T** [2007] Catecholaminergic neurotransmitters regulate migration and repopulation of immature human CD34+ cells through Wnt signaling. *Nat Immunol* 8: 1123-1131.
- [100] **Yang M, Li K, Ng PC, Chuen CK, Lau TK, Cheng YS, Liu YS, Li CK, Yuen PM, James AE, Lee SM and Fok TF** [2007] Promoting effects of serotonin on hematopoiesis: ex vivo expansion of cord blood CD34+ stem/progenitor cells, proliferation of bone marrow stromal cells, and antiapoptosis. *Stem Cells* 25: 1800-1806.
- [101] **Méndez-Ferrer S, Battista M and Frenette PS** [2010] Cooperation of β_2 - and β_3 -adrenergic receptors in hematopoietic progenitor cell mobilization. *Ann N Y Acad Sci* 1192: 139-144.

- [102] **Vasin MV, Ushakov IB, Semenova LA and Kovtun VIu** [2001] [Pharmacologic analysis of the radiation-protecting effect of indraline] *Radiats Biol Radioecol* 41: 307-309. Russian
- [103] **Maestroni GJM, Togni M and Covacci V** [1997] Norepinephrine protects mice from acute lethal doses of carboplatin. *Exp Hematol* 25: 491-494.
- [104] **Vasin MV, Ushakov IB, Kovtun VIu, Komarova SN and Semenova LA** [2006] Effect of radioprotector indralin on carboplatinum hemotoxicity. *Bull Exp Biol Med* 141: 437-439.
- [105] **Lissoni P** [2007] Biochemotherapy with immunomodulating pineal hormones other than melatonin: 5-methoxytryptamine as a new oncostatic pineal agent. *Pathol Biol (Paris)* 55: 198-200.
- [106] **Nefedova VV, Inzhevatin EV and Nefedov VP** [2002] Role of 5₂ receptors in the stimulatory effect of serotonin on hemopoietic bone marrow stem cells. *Bull Exp Biol Med* 133: 419-420.
- [107] **Muthu K, Iyer S, He L-K, Szilagyi A, Gamelli RL, Shankar R and Jones SB** [2007] Murine Hematopoietic Stem cells and Progenitors Express Adrenergic Receptors. *J Neuroimmunol* 186: 27-36.
- [108] **Chernov GA, Shliakova TG, Sharygin VL, Sharf VG, Todorov IN, Mitrokhin IuI, Efremova OI, Khristianovich DS, Rozantseva TV and Pulatova MK** [1994] [The molecular aspects of the action of the radioprotector indralin]. *Izv Akad Nauk Ser Biol* (1): 20-37. Russian
- [109] **Pulatova MK, Sharygin VL and Todorov IN** [1999] The activation of ribonucleotide reductase in animal organs as the cellular response against the treatment with DNA-damaging factors and the influence of radioprotectors on this effect. *Biochim Biophys Acta* 1453: 321-329
- [110] **Pulatova MK, Sharygin VL and Shliakova TG** [2003] [The reaction of deoxyribonucleotides synthesis on an irradiation and its modification by radioprotectants.] *Radiats Biol Radioecol* 43: 29-43. Russian
- [111] **Han J, Zou Z, Zhu C, Deng J, Wang J, Ran X, Shi C, Ai G, Li R, Cheng T and Su Y** [2009] DNA synthesis of rat bone marrow mesenchymal stem cells through alpha1-adrenergic receptors. *Arch Biochem Biophys* 490: 96-102.
- [112] **Ryan SV, Carrithers SL, Parkinson SJ, Skurk C, Nuss C, Pooler PM, Owen CS, Lefer AM and Waldman SA** [1996] Hypotensive mechanisms of amifostine. *J Clin Pharmacol* 36: 365-373.
- [113] **Romashko OO, Lebedev VG and Moroz BB** [1990] [The role of the hemopoietic microenvironment on the hemopoiesis-stimulating effect of cystamine]. *Radiobiologiya* 30: 779-784. Russian

Conflict of interest statement: The author declares no conflicts of interest and is alone responsible for the content and writing of this paper.